

Calix[6]arene derivative as chromogenic sensor for anti-hypertensive drugs

S K Menon*, P Jose, U Harikrishnan & U Pal

Department of Chemistry, School of Sciences, Gujarat University
Navrangpura, Ahmedabad 380 009, India
Email: sobhanamenon@rediffmail.com

Received 26 February 2007; revised 22 December 2007

A simple, rapid, and sensitive spectrophotometric method has been developed for the determination of atenolol, propranolol hydrochloride and metoprolol tartarate. The method is based on the reaction of these drugs as n-electron donors with acceptor groups on macrocyclic ring of calixarene. Due to the rapid development of color at ambient temperatures, the chromogenic calix[6]arene derivative can be used for the determination of these β -adrenergic blocking drugs. The association constants (K_c^{AD}) and free energies for the complexes have been determined. The proposed method can be used to determine the drugs in pharmaceutical tablets and urine.

IPC Code: Int. Cl.⁸ G01N21/00

Anti-hypertensive (β -adrenergic blocking) drugs, such as atenolol, propranolol hydrochloride, metoprolol tartarate, and methyl dopa are chemical agents that exert their principle pharmacological and therapeutic effects by acting at peripheral sites of the sympathetic nervous system¹. β -Adrenergic blocking drugs are used mainly in angina pectoris, certain arrhythmia, systematic hypertension, and other cardiovascular disorders such as arterial fibrillation, myocardial infarction, and sinus tachycardia².

Several methods have been described for the quantitative determination of the β -adrenergic blocking agents including spectrophotometric³⁻⁸, fluorimetric^{9,10} chromatographic¹¹⁻¹⁷ and electrochemical methods¹⁸⁻²¹. However, no method is reported using macrocyclic compounds like calixarenes, cyclodextrins, crown ethers, etc. These macrocyclic compounds have been extensively used to separate alkali, alkaline earth cations, heavy metals, and amines either by solvent extraction or by liquid membranes²²⁻²⁴.

We report herein a simple, sensitive, accurate and precise spectrophotometric method for the determination of atenolol, propranolol hydrochloride, metoprolol tartarate via reaction of these drugs with coumarin calix[6]arene hydroxamic acid (CoCHA). Calixarenes are popular host molecules for small

organic molecules. But till now, there have been very few reports of their inclusion complexes with drug molecules^{25,26}.

Experimental

Absorbance measurements were made on Hitachi 3210 UV-vis spectrophotometer with matched 10 mm quartz cells. pH measurements were made on Systronics pH-meter model 335 equipped with glass and calomel electrodes.

All solvents used were obtained from E. Merck (AR Grade) and used without further purification. Atenolol, propranolol hydrochloride and metoprolol tartarate were obtained as gift samples from Cadila Pharma, Ahmedabad. The standard stock was prepared by dissolving 200 mg of drug in 50 mL ethanol in a standard flask. Working solutions were subsequently prepared by appropriate dilution of the stock solution. CoCHA reagent solution (0.01%) solution was prepared in dichloromethane.

The nitro benzyl ester derivative of calix[6]arene was synthesized by stirring together methyl calix[6]arene (21.6 g, 0.03 mol) in dioxane(100 mL) and *p*-nitrobenzoyl chloride (5.6 g, 0.03 mol) in dichloromethane(20 ml) at 15-20°C for about 1 h. The yellow precipitate obtained was collected by filtration. Yield 12 g (75%).

For the partial reduction, nitro ester derivative of methyl calix [6] arene (12 g, 0.007 mol) was dissolved in dioxane:water (8:2, v/v); 4-5 g of Raney nickel (W-2) was added, followed by dropwise addition of hydrazine hydrate (80%) (20 mL, 0.41 mol). The reaction mixture was stirred below 0°C for 1 h, filtered and used directly for further synthesis. The partially reduced nitro ester derivative of calix[6]arene, thus prepared, was coupled directly with coumarin-3-carbonyl chloride to give the final compound, CoCHA. It was recrystallized using chloroform-heptane mixture. Yield 3 g (41%), m.pt.>300°C (decom.). (Found: C 68.25, H 4.03, N 4.30. Calc. for C₁₁₀H₇₈O₂₈N₆: C 68.39, H 4.04, N 4.35%). (KBr)/cm⁻¹ 3200, 1700, 1620, 1580, 1370. δ (CDCl₃) 10.3(2H, s, N-OH), 6.74-7.98 (46H, m, aromatic protons), 4.81 (4H, d, CH₂), 4.2 (4H, d, CH₂), 3.8 (2H, d, CH₂), 3.5 (2H, d, CH₂), 2.8 (6H, s, CH₃), 2.5 (12H, s, CH₃), *m/z* (FAB) 1930(M⁺).

An aliquot of the sample solution (0.5-20 μg of atenolol, 5-120 μg of propranolol hydrochloride and 2-150 μg of metoprolol tartarate) was transferred to 10 mL volumetric flask and mixed with 5 mL of the 0.01% reagent solution. pH of the solution was maintained at 5-6 using appropriate buffers. The colored solution was diluted to the mark with dichloromethane and measured at 440 nm, 427 nm, and 432 nm for atenolol, propranolol hydrochloride, and metoprolol tartarate, respectively, against the reagent blank.

A quantity of the powdered tablets equivalent to 50 mg of drug was dissolved in ethanol and filtered to remove the insoluble contents like binders and other excipients.

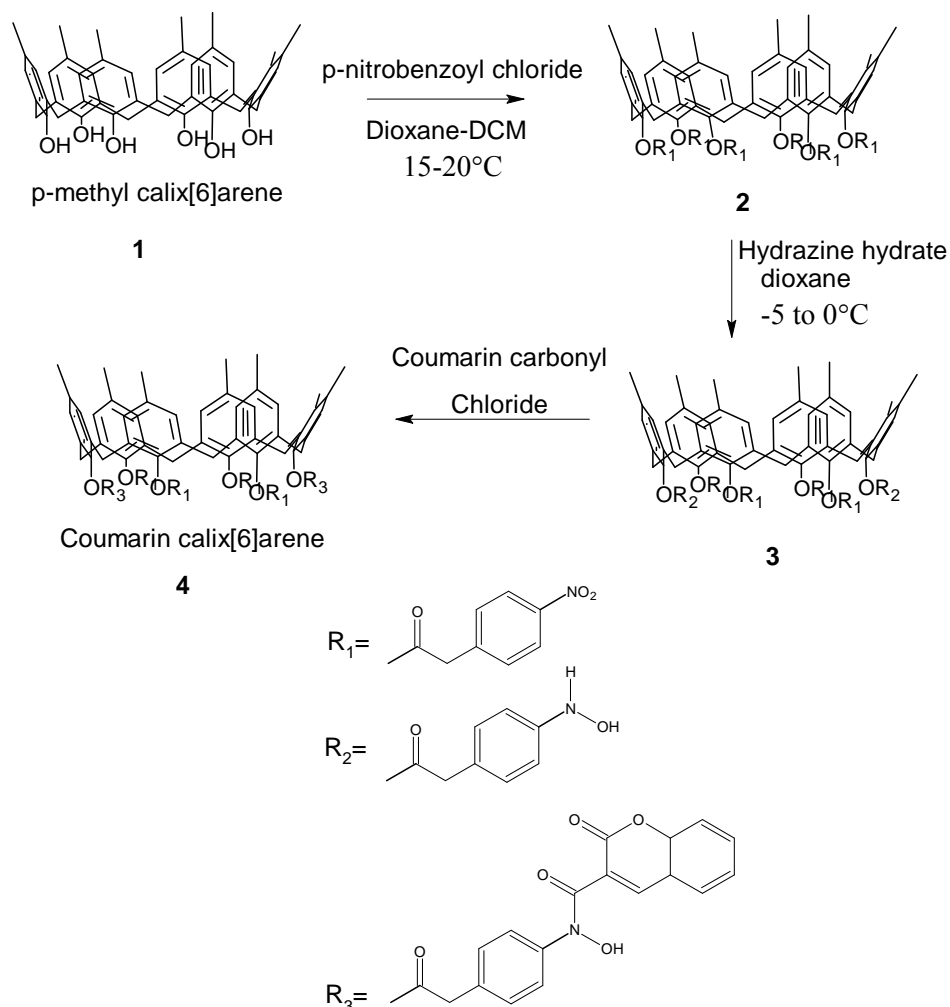
Urine samples were collected during 48 h after the oral intake of a 100 mg single dose of each drug.

Drug-free urine was also collected from two volunteers (male and female). Both types of samples were vortexed at 3800 rpm for 15 min and frozen until preparation for analysis. For the analysis of the urine samples, 10 mL of the sample was allowed to dry at ambient temperature and the residue was dissolved in ethanol for subsequent use.

Results and discussion

CoCHA was synthesized by the coupling of the partially reduced nitro ester derivative of methyl calix[6]arene with coumarin carbonyl chloride (Scheme 1). The synthesized compound CoCHA was characterized by its elemental analysis, FT-IR, ^1H NMR and FAB-MS.

The molecular interactions between electron donors and acceptors are generally associated with the



Scheme 1

Table 1—Comparative study of present method with other methods

Reagent	Molar absorptivity ($L \text{ mol}^{-1} \text{ cm}^{-1}$)		
	Atenolol	Propranolol	Metoprolol
Hydroxamic acid ³	5.3×10^2	-	-
Promethazine hydrochloride ³¹	-	1.36×10^4	-
Methdilazine hydrochloride ³¹	-	2.55×10^4	-
Chelation with Cu in CS_2 ³²	-	6.89×10^3	1.08×10^4
Chloranil ⁶	6.65×10^3	1.86×10^4	1.80×10^3
Iodine ⁶	3.75×10^3	6.89×10^3	7.12×10^3
Bromanil ⁶	1.05×10^4	1.04×10^4	1.36×10^4
Tetracyanoethylene ⁶	4.88×10^3	3.96×10^3	7.53×10^3
7,7,8,8-Tetracyanoquinodimethane ⁶	1.40×10^4	5.83×10^3	1.29×10^4
2,3-Dichloro-5,6-dicyano-1,4-benzoquinone ⁶	9.25×10^3	2.28×10^4	1.59×10^4
CoCHA (present method)	5.2×10^5	2.3×10^5	4.38×10^5

Table 2—Various parameters of the proposed method

Parameter	Atenolol	Propranolol	Metoprolol
Color developed	Red-orange	Yellow	Orange-yellow
Working pH	5-6	5-6	5-6
λ_{max} , nm	440	427	432
Beer's law range, $\mu\text{g/mL}$	0.05-2	0.5-12	0.2-15
Detection limit, $\mu\text{g/mL}$	0.023	0.18	0.076
Quantification limit, $\mu\text{g/mL}$	0.071	0.55	0.23
Molar absorptivity, $l \text{ mol}^{-1} \text{ cm}^{-1}$	5.2×10^5	2.3×10^5	4.38×10^5
Sandell sensitivity, $\mu\text{g cm}^{-2}$	0.0005	0.0001	0.0006
Regression equation ^a			
Intercept, <i>a</i>	- 0.0518	- 0.0089	- 0.0021
Slope, <i>b</i>	0.2035	0.0804	0.0788
Correlation coefficient	0.9990	0.9995	0.9991

^a $A = a + b \cdot C$, where *A* is absorbance and *C* is concentration in $\mu\text{g/mL}$

formation of intense colored charge transfer complexes, which absorb in the visible region. The photometric methods based on these interactions are usually simple and convenient because of the rapid formation of the complexes²⁷. β -Adrenergic drugs are good non-bonding electron donors and form charge transfer complexes with σ or π -acceptors. The interaction of all studied drugs with CoCHA in dichloromethane at room temperature gave colored charge-transfer complexes. There was a complete one-electron transfer from the drug donor to the acceptor CoCHA.

There was an immediate change of the pale yellow color of the reagent in dichloromethane (409 nm) to orange-red upon addition of atenolol (440 nm), propranolol hydrochloride (427 nm) or metoprolol tartarate (432 nm). The π - π stacking interactions due to the aromatic rings attached to the drug molecules and CoCHA, and the strong hydrogen bonding by the hydroxamic acid moiety provide stability to the complex. A comparative study of this method with

Table 3—Evaluation of the accuracy and precision of the proposed method

Drugs	Conc. ($\mu\text{g/mL}$)	Recovery (%)	RSD ^a (%)	Lit. method ²⁸
Atenolol	0.5	100.10	0.90	100.20
	1.0	98.90	0.89	99.90
	1.5	99.00	0.94	100.10
	2.0	100.14	0.91	100.04
Propranolol	1.0	99.96	0.87	100.00
	2.0	98.99	0.88	98.97
	3.0	100.01	0.90	100.03
	4.0	99.90	0.84	99.96
Metoprolol	2.0	99.02	0.95	100.02
	4.0	99.03	0.94	99.09
	6.0	99.01	0.92	100.00
	8.0	100.14	0.89	100.09

^aAverage of six determinations

Table 4—Physical parameters of the proposed method

Drug	Association constant/ formation constant	Free energy ΔG (kJ/mol)
Atenolol	3.8×10^2	-15.0
Propranolol	3.4×10^4	-32.05
Metoprolol	3.8×10^3	-26.48

Table 5—Analysis of the formulations

Drug	Preparation	Nominal value (mg/tablet)	Recovery (%)	RSD ^a (%)	Lit. method ²⁸
Atenolol	Tenormin (Nicholas Piramal)	50	99.0	0.89	100.14
	Hipres-50 (Cipla)	50	98.5	0.78	98.00
	Coronol 50 (Sarabhai Chemicals)	50	99.0	0.43	99.30
Propranolol	Ciplar LA-40 (Cipla)	40	99.9	0.89	100.08
	Inderal (Nicholas Piramal)	40	101.0	0.86	101.80
	Corbeta Sarabhai (Chemicals)	40	100.2	0.58	100.3
Metoprolol	Metolar -H (Cipla)	50	100.8	0.12	101.0
	Betaloc (Astra Zeneca)	50	100.1	0.91	100.6
	Lopresor (Novartis)	50	99.9	0.89	100.0

^a = average of six determinations; RSD=Relative standard deviation

Table 6—Analysis of urine samples

Drug	Sample	Amt. (mg)	RSD ^b (%)	Lit. method ²⁸
Atenolol	1	1.6	1.5	1.7
	2	2.8	0.6	2.8
	3	2.9	0.9	2.9
	4	1.8	0.6	1.9
	5	2.9	0.7	2.9
Propranolol	1	3.6	1.5	3.7
	2	3.8	0.6	3.8
	3	3.9	0.5	3.9
	4	3.8	0.6	3.8
	5	2.9	0.9	2.9
Metoprolol	1	2.6	1.5	2.7
	2	2.8	0.8	2.8
	3	1.9	0.7	1.9
	4	2.8	0.6	3.8
	5	2.9	0.9	3.9

^bAverage of six determinations

other reported methods is as shown in Table 1. All assays in the proposed method are performed in the visible region and do not have interferences from extracted excipients of dosage forms.

The complexation of the anti-hypertensive drugs was studied under the optimum conditions of pH, nature of solvent, and reagent concentration. Optimum pH range for complexation was found to be 5.0-6.0. Beyond this pH range, complexation of the drugs was incomplete. Common organic solvents like chloroform, dichloromethane, ethanol and carbon tetrachloride were studied. But, optimum absorbance was found when dichloromethane was used.

The amount of reagent needed to obtain maximum and reproducible absorbance was 5 mL for atenolol,

7 mL for propranolol and 5 mL for metoprolol tartarate.

The general spectral characteristics like Beer's law range, molar absorptivity, Sandell's sensitivity, regression equations, and correlation coefficients obtained by the linear treatment of the results are given in Table 2. The limit of detection and limit of quantification were calculated from the standard deviations of the absorbance measurements obtained from a series of 13 blank solutions for each procedure.

In order to compare the accuracy and precision of the proposed method with the official method, six replicate analyses were performed on four standards. The values obtained for the students t-test and the variance ratio F-test are less than the critical values suggesting that the proposed method is precise and accurate as the USP methods²⁸ (Table 3).

Interference studies

In order to examine the selectivity of the proposed method to pharmaceutical preparations and urine samples, effect of common additives, adjuvants and excipients (glucose, lactose, talc, magnesium stearate and starch) as well as albumin (excreted in the urine) was experimentally studied. An error of not more than $\pm 3\%$ in the absorbance was taken as interference. The results show that the common additives and excipients cause no interference in the proposed method.

Physical parameters

The association constants were calculated for the interaction of each drug with CoCHA using Benesi-Hildebrand equation²⁹.

The standard free energy changes of complexation (ΔG°) were calculated from the association constants³⁰.

The association constants and the free energy reveal that the complexes formed with the drugs are stable (Table 4). The high values of association constants are common in n-electron donors where the intermolecular overlap may be considerable.

Analysis of pharmaceutical dosage forms and urine samples

The results were compared statistically with those obtained by applying the literature method²⁸ (Tables 5 and 6). The percentage recoveries were in the range of 98-100 implying that the method can be satisfactorily used for the spectrophotometric determination of the studied drugs.

Acknowledgement

The authors wish to thank Gujarat Council of Science and Technology (GUJCOST/200531/CE/03-04) and UGC, New Delhi (No.F/540/5/DRS/2002 SAP-I) for financial assistance and CDRI, Lucknow for spectral data.

References

- Delgado J N & Remers W A, *Wilson and Gisvold's Textbook of Organic Medicinal and Pharmaceutical Chemistry*, 9th Edn (JP Lippincot Company, New York), 1991, pp. 413.
- Andrejus K, *Essentials of Medicinal Chemistry*, 2nd Edn (John Wiley & Sons Inc, USA) 1988.
- Agrawal Y K, Menon S K, Rajput S & Raman K, *Anal Lett*, 25 (1992) 1503.
- Alpdogan G & Sungur S, *Spectrochim Acta*, 55A (1999) 2705.
- Satuf M L, Robles J C, Goicoechea H C & Olivieri A C, *Anal Lett*, 32 (1999) 2019.
- Salem H, *J Pharm Biomed Anal*, 29 (2002) 527.
- Golcu A, Yucesoy C & Serin S, *Il Farmaco*, 59 (2004) 487.
- Al-Ghannam S M, *J Pharm Biomed Anal*, 40 (2006) 151.
- Gajewska M, Glass G & Kostelecki, *J Acta Pol Pharm*, 49 (1992) 1.
- Yang M, Zhang W W & Wang H N, *Fenxi Huaxue*, 24 (1996) 740.
- Veronico M, Rango G & Vetuschi C, *Spec Lett*, 28 (1995) 60.
- Lukkari P & Siren H, *J Chromatogr A*, 717 (1995) 58.
- Chiap P, Hubert P, Boulanger P & Crommen J, *Anal Chim Acta*, 391 (1999) 227.
- Wy J C, Lord H L, Pawliszyn J & Kataoka H, *J Microcol Sep*, 12 (2000) 255.
- Gillott N C, Euerby M R, Johnson C M, Barrett D A & Shaw P N, *Chromatographia*, 51 (2000) 167.
- Svensson S, Karlsson A, Gyllenhaal O & Vessman J, *Chromatographia*, 51 (2000) 283.
- Cheng C & Chou C *Chromatogr. J Biomed Appl*, 762 (2001) 51.
- Alvarez-Lueje A, Nunez-Verqara L J & Sequella J A, *Il Farmaco*, 46 (1991) 593.
- Issa Y M & Amin A S, *Mikrochim Acta*, 118 (1995) 85.
- Maguregui M I & Alonoso R M, *Chromat J Biomed Appl*, 674 (1995) 85.
- Billiot E & Warner I M, *Anal Chem*, 72 (2000) 1740.
- Gutsche C D, *Calixarenes*, 1st Edn (Wiley, London), 1989.
- Takeshita M & Shinkai S, *Bull Chem Soc Japan*, 68 (1995) 1088.
- Bohmer V, *Angew Chem Int Ed Engl*, 34 (1995) 713.
- Tung C & Ji H, *J Chem Soc Perkin Trans 2*, (1997) 185.
- Yang W & de Villiers M M, *Eur J Pharm Biopharm*, 58 (2004) 629.
- Foster R, *Organic Charge Transfer Complexes*, 1st Edn (Academic Press, London) 1969.
- The United States Pharmacopoeia 24, National Formulary*, US Pharmacopoeial Convention, Rockville, MD, 2000, pp. 19.
- Benesi H A & Hildebrand J, *J Am Chem Soc*, 71 (1949) 2703.
- Martin A N, Warbrick J S & Cammarata A, *Physical Pharmacy*, 3rd Edn (Lea & Feibiger Press, Philadelphia) 1969.
- Gowda B G, Seetharamappa J & Melwani M B, *Anal Sci*, 8 (2002) 671.
- El-Ries M A, Attia Abou F M & Ibrahim S A, *J Pharm Biomed Anal*, 15 (2000) 179.