

Toxicity of cypermethrin to the juveniles of freshwater fish *Poecilia reticulata* (Peters) in relation to selected environmental variables

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Abstract

Toxicity of cypermethrin, a synthetic pyrethroid insecticide, was evaluated for the juveniles (8.5 ± 1.5 mm) of a freshwater fish, *Poecilia reticulata* (Peters) in relation to selected environmental variables such as temperature, hardness, pH and salinity. The LC_{50} 's were found to change significantly with the change in temperature, hardness, pH and salinity of water. The juveniles were found more susceptible to cypermethrin at low water hardness (270 ± 1 mg/l) and low water pH (5.4) as compared to other variables. Based on LC_{50} 's the order of toxicity of cypermethrin to juveniles of *Poecilia* was observed as: pH < salinity < hardness < temperature. The range of safe dischargeable concentrations of cypermethrin (1.04-1.09 ppb) was too low as compared to harmless or safe concentrations (45.18-75.25 ppb) for the juveniles at selected environmental variables.

Keywords: Toxicity, Cypermethrin, Freshwater fish, *Poecilia reticulata*, Environmental variables.

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Poecilia reticulata

and $25 \pm 1^\circ\text{C}$), hardness (270 ± 1 and 660 ± 5 mg/l), pH (5.4, 7.6 and 9.1) and water salinities (0.53, 1.18 and 1.54%) following standard methods of APHA⁹. The water having different hardness values, i.e. 270 ± 1 and 660 ± 5 mg/l were collected from public supply and bore well, respectively. The water of different pH, i.e. 5.4 and 9.1 were prepared by adding HCl and NaOH, respectively. Also the water of different salinity, i.e. 1.18 and 1.54% were prepared by adding common salt (NaCl). The pH and salinity of normal water were estimated as 7.6 and 0.53%, respectively. The experimental water were analysed for selected physico-chemical characteristics as per standard methods of APHA⁹. Glass jars of one litre capacity were used for conducting static bioassay tests. The juveniles (8.5 ± 1.5 mm) produced from healthy males and females of *Poecilia* were maintained in 100 litre capacity of plastic tank on artificial diet (rice bran and ground nut cake mixture - 1:1).

Introduction

Cypermethrin (pyrethroid) is a widely used synthetic insecticide and has been found highly toxic for fish populations and aquatic invertebrates¹⁻⁴. It is used in cotton, cereals, vegetables, fruits, food storage and in animal husbandry. Reports are also available on the toxicity of cypermethrin on the early stages of fish⁵⁻⁷. Further, activities of enzymes such as Glutamic oxaloacetic acid transaminase (GOT), Glutamic pyruvic acid transaminase (GPT) and Lactate dehydrogenase (LDH) have also changed marginally due to cumulative effects of cypermethrin in heart, liver and kidney of *Cyprinus carpio* Linn.⁸ However, reports on cypermethrin toxicity

in relation to environmental variable on early stages of fish are not available. In view of this, an attempt has been made to assess the effects of selected environmental variables, viz. temperature, hardness, pH and salinity on the short-term (96 h) toxicity of cypermethrin to the juveniles of a freshwater fish *Poecilia reticulata* (Peters) considering various parameters such as median lethal concentration (LC_{50}), safe or harmless concentration and safe dischargeable concentration.

Materials and Methods

Short-term (96 h) static bioassays for cypermethrin to juveniles were conducted at selected environmental variables such as temperatures (16.5 ± 1

The pesticide cypermethrin marketed and manufactured by Crystal Phosphate Limited (jakpot 25 EC) was used in the present investigation. To commence with assays for cypermethrin a common stock solution was prepared by dissolving appropriate amount in one litre of deionized diluent water as per the following formula: $N_1V_1 = N_2V_2$ [where N_1 -concentration of commercial formulation in per cent, V_1 -volume or amount of commercial formulation required in ml, N_2 -desired concentration of formulation required in ml, V_2 - volume or amount of solution required in ml]. The series of different nominal concentrations of cypermethrin (as ppb) based on the progressive bisection of intervals on a logarithmic scale were prepared by adding stock solution into the measured diluent's water⁹.

The experimental units, i.e. 1 litre l glass jars were filled with toxicant solutions and placed in three rows. Full scale experiments were conducted in triplicate by testing various concentrations

in the range known by preliminary exploratory test. Each glass jar was labelled with the details such as concentration, water quality variables, replicate number, date and time of the experiment. Ten acclimatized healthy juveniles of size (8.5 ± 1.5 mm) were transferred to these jars after about 20 minutes of the preparation of test solution. Proper controls were run simultaneously. The experiments were continued for a period of 96h. The number of dead juveniles in each concentration of cypermethrin were observed carefully and recorded at the time intervals of 24, 48, 72 and 96h. The LC_{50} 's of cypermethrin for juveniles in relation to each environmental variable were estimated for the time intervals of 24, 48, 72 and 96h by probit analysis¹⁰. Presumable safe or harmless and safe dischargeable concentrations of cypermethrin were calculated by using the formula of Hart *et al*¹¹.

Results and Discussion

The results revealed that cypermethrin is highly toxic to the juveniles of *Poecilia* as the LC_{50} values were noticed in ppb. The LC_{50} 's were also found to change significantly with the change in temperature, hardness, pH and salinity (Table 1). The safe concentrations of cypermethrin for the juveniles were different for each selected environmental variables. The bioassays conducted at water of higher hardness (660 ± 5 mg/l), moderate pH (7.6) and normal salinity (0.53%) at a temperature of $16.5 \pm 1^\circ\text{C}$ showed slightly higher values of harmless concentrations (75.25 ppb) as compared to that of bioassays conducted at higher temperature ($25 \pm 1^\circ\text{C}$; 56.10 ppb), hardness (270 ± 1 mg/l; 45.62 ppb), both acidic and alkaline pH (5.4 and 9.1; 45.18 and 47.45ppb) and higher salinity levels (1.18 and 1.54%; 57.29 and 50.31 ppb). The calculated safe dischargeable concentrations of cypermethrin were recorded too low as compared to safe harmless concentrations (Table 1).

Table 1 : Toxicity of cypermethrin for the juveniles of *Poecilia reticulata* (Peters) in relation to temperature, hardness, pH and salinity

Environmental variables	Durations (h)	$LC_{50, s}$ (ppb)	95% confidence limit (UCL-LCL)	R = UCL-LCL	Heterogeneity factor	Safe or harmless concentration (as ppb)	Safe dischargeable concentrations (as ppb)
Temperature a. $16.5 \pm 1^\circ\text{C}$	24	316.59	478.10-209.64	2.28	0.22	75.25	1.06
	48	281.71	351.13-226.01	1.55	0.25		
	72	248.96	309.81-200.07	1.54	0.31		
	96	221.96	264.03-186.60	1.41	0.30		
b. $25 \pm 1^\circ\text{C}$	24	219.83	240.86-200.64	1.20	25.82	56.10	1.04
	48	202.72	228.37-179.95	1.26	0.75		
	72	186.22	210.93-164.41	1.28	0.28		
	96	168.07	191.94-147.16	1.30	0.44		

Environmental variables	Durations (h)	LC _{50, s} (ppb)	95% confidence limit (UCL-LCL)	R = UCL-LCL	Heterogeneity factor	Safe or harmless concentration (as ppb)	Safe dischargeable concentrations (as ppb)	
Hardness a. 270±1 mg/l	24	184.06	200.23-169.19	1.18	17.23	45.62	1.05	
	48	167.28	189.45-147.71	1.28	0.05			
	72	155.55	171.59-141.01	1.21	0.06			
	96	148.80	161.91-136.75	1.18	0.66			
	b. 660±5 mg/l	24	316.59	478.10-209.64	2.28	0.22	75.25	1.06
		48	281.71	351.13-226.01	1.55	0.25		
		72	248.96	309.81-200.07	1.54	0.31		
		96	221.96	264.03-186.60	1.41	0.30		
pH	a. 5.4	24	179.65	200.18-161.23	1.24	0.82	45.18	1.04
		48	164.47	180.93-149.51	1.21	4.02		
		72	160.64	181.02-142.55	1.26	0.78		
		96	148.89	171.33-124.40	1.32	0.86		
	b. 7.6	24	316.59	478.10-209.69	2.28	0.22	75.25	1.06
		48	281.71	351.13-226.01	1.55	0.25		
		72	248.96	309.81-200.07	1.54	0.31		
		96	221.96	264.03-186.60	1.41	0.30		
	c. 9.1	24	190.47	215.47-168.37	1.27	0.29	47.45	1.05
		48	173.53	195.43-154.09	1.26	0.39		
		72	160.63	179.13-144.03	1.24	5.71		
		96	153.34	178.30-131.86	1.35	5.23		
Salinity	a. 0.53%	24	316.59	478.10-209.64	2.28	0.22	75.25	1.06
		48	281.71	351.13-226.01	1.55	0.25		
		72	248.96	309.81-200.07	1.54	0.31		
		96	221.96	264.03-186.60	1.41	0.30		
	b. 1.18%	24	241.78	338.29-172.80	1.95	0.08	57.29	1.06
		48	214.85	265.36-173.95	1.52	0.36		
		72	188.59	235.91-150.77	1.56	0.12		
		96	168.37	238.03-119.09	1.99	0.07		
	c. 1.54%	24	236.31	407.98-136.87	2.98	0.06	50.31	1.09
		48	199.06	249.02-159.12	1.56	0.17		
		72	184.03	212.59-159.30	1.33	0.64		
		96	161.61	179.32-145.65	1.23	5.15		

LC_{50s} - Median lethal concentrations; UCL - Upper confidence limit; LCL - Lower confidence limit; R- Ratio of confidence limits

Characteristics of the experimental water revealed that they were similar to the standard values under natural conditions as they did not contain any toxic substances. However, few changes have been observed in some of the characteristics of water such as hardness, total alkalinity, dissolved oxygen and electrical conductivity (Table 2).

Polat *et al*⁶ have also investigated the acute toxicity of beta-cypermethrin and estimated 48h LC₅₀ value for guppy (*P. reticulata*) as 21.4 µg/l at a temperature of 22 ± 1°C. Whereas, Aguigwo⁵ has recorded much higher 96h LC₅₀'s value of cypermethrin as 4.17 mg/l for *Clarias gariepinus* and also found that mortality increased with increase in concentrations. They however, observed that number of dead embryos of the common carp (*C. carpio*) significantly increased in response to even very low cypermethrin concentrations of 0.0001, 0.001, 0.01, 0.1, 1, 2, 4 and 8 µg/l and estimated the 48 h LC₅₀'s (with 95%

confidence limits) and 96h LC₅₀'s (with 95% confidence limits) of cypermethrin for common carp embryos and larvae as 0.909 (0.256-5.074) and 0.809 (0.530-1.308) µg/l, respectively⁷. Wang *et al*⁴ determined the LC₅₀ of cypermethrin at 96h for carp as 12.6µg/l. However, Collins and Cappello³ have recorded the LC₅₀'s of cypermethrin to freshwater prawn *Palaemonetes argentinus* as 0.0031 and 0.0020µg/l for 24 and 96h, respectively. The present investigation is in agreement with the above findings as the developing stage, viz. juveniles of *Poecilia* showed highly susceptibility to even very low concentrations of cypermethrin at all selected environmental variables. The results of the present study also revealed that changes in the selected water quality parameters, viz. temperature, hardness, pH and salinity played significant role in intoxication of cypermethrin to the juveniles of *Poecilia* as considerable difference in LC₅₀'s was noticed for different durations. Vardia and

Durve¹² also found 7-7.5 fold increase in toxicity of 2,4-D to *C. carpio* with the rise in temperature from 17°C (Winter) to 39°C (Summer). Whereas Baer *et al*¹³ have noticed that the 96h LC₅₀ of profenophos to fathead minnows (*Pimephales promelas*) at higher temperature of 30 ± 2°C was significantly lower as 21.5µg/l as compared to 333µg/l at a lower temperature of 20 ± 2°C. More or less similar results have been observed in the present study as the higher 96h-LC₅₀ (221.96ppb) of cypermethrin to the juveniles was observed at a temperature of 16.5±1°C as compared to lower 96h LC₅₀ (168.07 ppb) at a temperature of 25 ± 1°C. Cairns *et al*¹⁴ also opined that temperature influences toxicity of chemicals to aquatic organisms. Whereas, Hashimoto and Nishiuchi¹⁵ have reported that the majority of the pesticides are more toxic to carps and daphnids at higher temperatures, but some including folpet and DDT are less toxic at higher temperatures.

Table 2 : Physico-chemical characteristics of the water used in static bioassays

Characteristics	Borewell water		Public supply water	With change in pH of water		With change in salinity of water	
	Winter	Summer		A	B	A	B
Water temperature (°C)	16.5±1	25±1	16.5±1	16.5±1	16.5±1	16.5±1	16.5±1
pH	7.6	7.7	7.8	9.1	5.4	7.9	8.0
Dissolved oxygen (mg/l)	6.5	5.2	6.4	6.2	6.1	5.8	5.6
Total alkalinity (mg/l)	600	540	340	420	380	500	450
Hardness (mg/l)	660±5	655±5	270±1	665±5	657±5	662±5	666±5
Salinity (%)	0.53	0.54	0.4	0.54	0.51	1.18	1.54
Electrical conductivity (µS)	2600	2700	1600	4800	5000	5400	6300

Values are S.E.; n=10

The present results are in agreement with the above findings as low LC_{50} 's of cypermethrin for the juveniles were recorded at a higher temperature of $25 \pm 1^\circ C$. Further, results of the present study also revealed that hardness of water plays a significant role in determining the toxicity of cypermethrin to juveniles of *Poecilia*. According to McCann and Hitch¹⁶ the toxicity of aquazine has been found to be highly significant in both soft and hard water to striped bass (*Morone saxatilis*) and reported a common 96h LC_{50} as 180 mg/l. In the present study sensitivity of the juveniles of *Poecilia* to cypermethrin was decreased considerably with the increase in hardness of water. Datta *et al*¹⁷ observed that the hardness of water significantly reduced the toxicity of the pesticide deltamethrin to scale carp fry (*C. carpio* var. *communis*). According to them the 96h LC_{50} values of deltamethrin, which was observed as 0.102 mg/l in soft water, increased to 0.8 mg/l in hard water. Further, Donald *et al*¹⁸ have observed that decrease in water hardness associated with decrease in alkalinity and pH also increased the toxicity of mixture of inorganic and organic contaminants. The same results have been obtained in the present study as the toxicity of cypermethrin to juveniles decreased subsequently with the increase in water hardness. Khillare and Wagh¹⁹ have reported that LC_{50} values of endosulfan 35 EC for 96h increases with pH and alkalinity. The present results are also in agreement with the findings of Wang *et al*²⁰, who suggested that salinity of water significantly enhanced aldicarb toxicity, cholinesterase inhibition and 14C aldicarb sulfoxide formation in rainbow

trout (*Oncorhynchus mykiss*). Abir *et al*²¹ have reported that environmental factors, such as salinity in addition to gender and development have significant impacts on the acute toxicity of aldicarb to Japanese medaka (*Oryzias latipes*). They also found that after 48h of aldicarb exposure, mortality of Japanese medaka was significantly increased ($P < 0.05$) in males from $13 \pm 5.7\%$ at 1.5 % salinity to $56 \pm 5.7\%$ at 20% salinity. In females also the mortality significantly increased ($P < 0.01$) from 17 ± 5.7 to $76 \pm 5.6\%$ at the same water salinity. Further, Evelyn *et al*²² have also reported that the toxicity of organophosphorus insecticides increased with higher salinity.

In the present investigation, range of presumable safe or harmless concentrations and dischargeable concentrations of cypermethrin for the juveniles are interesting and significant since they are not constant with the succeeding experiments conducted for the selected environmental variables. More or less similar pattern of safe concentrations have been reported by other workers^{7, 23-25}.

Conclusion

The result suggests that cypermethrin is highly toxic to the juveniles of *P. reticulata* at selected environmental variables (pH < salinity < hardness < temperature). It is also recommended that before using cypermethrin in any aquaculture processes, the estimated safe and dischargeable concentrations should be considered important to protect living organisms as well as fish.

References

1. Gowland BTG, Moffat CF, Stagg RM, Hovlioh DF and Davies IM, Cypermethrin induces glutathione S-transferase activity in the shore crab, *Carcinus maenas*, *Marine Environ Res*, 2002, **54**, 169-177.
2. Sarkar B, Chatterjee A, Adhikari S and Ayyappan S, Carbofuran and cypermethrin induced histopathological alterations in the liver of *Labeo rohita* (Ham) and its recovery, *J Appl Ichthyol*, 2005, **21**, 131.
3. Collins P and Cappello S, Cypermethrin toxicity to aquatic life: Bioassay for the freshwater prawn, *Palaeomonetes argentinus*, *Arch Environ Contam Toxicol*, 2006, **18**, 10-14.
4. Wang Y, Xiong L, Liu XP, Xie T, Wang K, Huang XQ and Feng ZL, Subacute toxicity of cypermethrin to carp, *J Agro-Environ Sci*, 2006, **25**, 200-203.
5. Aguigwo JN, The toxic effect of cymbush pesticide on growth and survival of African catfish, *Clarias gariepinus* (Burchell), *J Aqua Sci*, 2002, **17**, 81-84.
6. Polat H, Erkoç FU, Viran R and Kocak O, Investigation of acute toxicity of beta-cypermethrin on guppies *Poecilia reticulata*, *Chemosphere*, 2002, **49**, 39-44.
7. Rahmi A, Kenan K, Mustafa D, Sibel SK and Murat P, Acute toxicity of synthetic pyrethroid cypermethrin on the common carp (*Cyprinus carpio* L.) embryos and larvae, *Aquacul Int*, 2005, **13**, 205.
8. Sivakumari K, Manavalaramanujam R, Ramesh M and Lakshmi R, Cypermethrin toxicity: sublethal effect of enzyme activities in a freshwater fish, *Cyprinus carpio* var. *communis*, *J Environ Biol*, 1997, **18**, 121-125.
9. American Public Health Association (APHA): American Water Works Association (AWWA)

- and Water Pollution Control Federation (WPCF), Standard Methods for the Examination of Water and Wastewater, 21st Edn, 2005, APHA, Washington DC.
10. Finney DJ, Probit Analysis, University Press, Cambridge, 1971, 333 pp.
 11. Hart WB, Doudoroff P and Greenbank J, The evaluation of the toxicity of industrial wastes, chemical and other substances to freshwater fishes, Atlant Refinding Co. (Phill), 1945, 317 pp.
 12. Vardia HK and Durve VS, The toxicity of 2, 4-D to *Cyprinus carpio* var. *communis* in relation to the seasonal variation in the temperature, *Hydrobiology*, 1981, **77**, 148-152.
 13. Baer KN, Olivier K and Pope CN, Influence of temperature and dissolved oxygen on the acute toxicity of profenofos to fathead minnows (*Pimephales promelas*), *Drug Chem Toxicol*, 2002, **25**, 231-245.
 14. Cairns J Jr, Heath AG and Parkar DC, The effects of temperature upon the toxicity of chemicals to aquatic organisms, *Hydrobiology*, 1975, **47**, 135-171.
 15. Hashimoto Y and Nishiuchi Y, Establishment of bioassay methods for the evaluation of acute toxicity of pesticides to aquatic organisms, *J Pest Sci*, 1981, **6**, 257-264.
 16. McCann JP and Hitch RK, Simazine toxicity to fingerling striped bass, *Progressive Fish Culturist*, 1980, **42**, 180-181.
 17. Datta S, Shah PS and Das RC, Soil sediment and hardness of water reduce the acute toxicity of deltamethrin to scale carp, *Pest Res J*, 2002, **14**, 327-336.
 18. Donald P, Joseph BH and James FD, Sensitivity of young striped bass to organic and inorganic contaminants in fresh and saline waters, *Trans Amer Fish Soc*, 2002, **114**, 748-753.
 19. Khillare YK and Wagh SB, Acute toxicity of the pesticide endosulfan to fishes, *Environ Ecol*, 1987, **5**, 804-806.
 20. Wang J, Girsle S and Schlenk D, Effects of a salinity on aldicarb toxicity in juvenile rainbow trout (*Oncorhynchus mykiss*) and striped bass (*Morone saxatilis* x *chrysops*), *Science*, 2001, **64**, 200-207.
 21. Abir T, Alfay E, Grisle S and Schlenk D, Characterization of salinity enhanced toxicity of aldicarb to Japanese Medaka: Sexual and developmental differences, *Environ Toxicol Chem*, 2001, **20**, 2093-2098.
 22. Evelyn H, Heugens W, Hendricks AJ, Dekkor T, Straalen NM Van and Admiraal W, A review of the effects of multiple stressors on aquatic organisms and analysis of uncertainty factors for use in risk assessment, *Crit Rev Toxicol*, 2002, **31**, 247-284.
 23. Srivastava SK, Jaiswal R and Srivastav AK, Lethal toxicity of deltamethrin (Decis) to a freshwater fish, *Heteropneustes fossilis*, *J Adv Zool*, 1997, **18**, 23-26.
 24. Ural MS and Saglam N, A study on the acute toxicity of pyrethroid deltamethrin on the fry rainbow trout (*Oncorhynchus mykiss*), *Pest Biochem Physiol*, 2005, **83**, 124-131.
 25. Koprucu SS, Koprucu K, Ural MS, Ispir U and Pala M, Acute toxicity of organophosphorus pesticide diazinon and its effects on behaviour and some hematological parameters of fingerling European cat fish (*Silurus glanis* L.), *Pest Biochem Physiol*, 2006, **86**, 96-105.