

Effect of exposure to extremely low electro-magnetic field during prenatal period on mice spleen

Parvin Dokht Bayat¹, Ali Ghanbari^{2*}, Saeid Babaei¹, Mozafar Khazaei², Rostam Ghorbani² & Mahmood Ayubian³

¹Department of Anatomy, Arak University of Medical Sciences, Arak, Iran

²Fertility and Infertility Research Center, Kermanshah University of Medical Sciences, P.O. Box 1568, Kermanshah, Iran

³Department of Anatomy, Tehran University of Medical Sciences, Tehran, Iran

Received 8 November 2010; revised 5 May 2011

Total body weight of newborns, the volume of spleen, and the number of megakaryocytes decreased following the exposure to ELF-MF (6×10^{-3} T and 50 Hz) at 1-5, 6-10, 11-15, and 16-20 days of pregnancy of mice. The complete period of gestation was sensitive to ELF-MF exposure; the initial days were more prone to exposure. The results suggest that the use of ELF-MF producing instruments should be limited during gestation.

Keywords: Electromagnetic field, Megakaryocyte, Mice, Spleen, Stereology

The overuse of cell phone and other instruments that generate extremely low electromagnetic fields (ELF-MF) made researchers interested in exploring the biological effects of these fields. Epidemiologically, the effects of these fields on embryogenesis have suggested a possible relationship between ELF-MF and adverse pregnancy outcomes in humans¹⁻³.

In vivo studies have suggested that ELF-MF disturb the development of non-mammalian embryos leading to induction of structural anomalies^{4,5} and markedly depress levels of cellular products⁶. In mammals results have been inconsistent, some studies reported no or only slight effects on fetuses that included slight limb anomalies and body weight reduction⁷⁻⁹. But others have shown that prenatal exposure to these fields induces adverse effects such as growth retardation, delayed puberty and reduced IGF-1 levels, damage of hypothalamus, pituitary gland and ovaries in female newborn Wistar rats¹⁰. Moreover, only 48 h exposure to these fields in pregnant rats increases incidence of non-vital fetuses, neonates, or infants¹¹. At cellular level many studies have shown that both prenatal and post-natal exposure to these fields induces noxious effects in mammals^{10,12-15}.

It seems that the effects of electromagnetic fields on embryonic development differ based on the type of species, parameters of the field and the type of tissue. Thus, to achieve more correct data, the more numerical modeling provides the most suitable analysis tool for more accurate estimation¹⁶. Two recent studies carried out stereological technique to evaluate quantitative changes induced by these fields^{13,14}.

The present study has been undertaken to see the effects of ELF-MF on the embryogenesis of mammals particularly on the development of mouse spleen. Stereological method has been used to evaluate the quantitative effects of ELF-MF on the volume of spleen and the number of megakaryocytes.

Materials and Methods

Forty-eight female NMRI (Naval Medical Research Institute) mice (10 weeks of age, 30 ± 4 g) purchased from Razi Institute (Karaj, Iran) were maintained in the animal house at Arak University of Medical Sciences. All animals received care as recommended by the research committee of the University. The mice were maintained on a regular diet and water at a 12:12 h L:D cycle without any stressful stimuli. at $23 \pm 2^\circ\text{C}$. Experiments were started after one week adaptation. Following mating, the beginning of pregnancy was determined by daily evaluation of sperm presence in vaginal smear by

*Correspondent author
Telephone: +98-831-4274617
Fax: +98-831-4281563
E-mail: aghanbari@kums.ac.ir

light microscopy analysis (ZEISS, magnification $\times 100$). Fertilized mice were divided randomly into following 6 groups of 8 mice in each. Group 1 (control): mice were placed in switch off electromagnetic housing while group 2 (sham): mice had no treatment at all. In the four experimental (E) groups pregnant mice were exposed to an established pulsed ELF-MF exposure system (6 mT and 50 Hz for 10 h a day for 5 days) at 1-5 (E1), 6-10 (E2), 11-15 (E3), and 16-20 (E4) days of gestation.

Pulsed ELF-MF exposure system

The ELF-MF used in the present study was produced by a pair of Helmholtz coils able to generate a highly homogeneous field (with homogeneity 5/1000). The characteristics of the system were as follows:

- (i) Power supply: 220 V in, 25 V out, permanent current intensity 3 Ampere.
- (ii) Multi-meter to control the intensity of the current entering the instrument.
- (iii) A 50 Hz sinusoidal oscillating ELF-MF was produced by a 380 round turn coil twisted around a cylinder (19 cm diameter and 15.5 cm length) and containing a chamber to house mice in the center of the cylinder, where the maximum even ELF-MF (6 mT) was recorded.
- (iv) A Teslometer (Compensation-51662) for precise measurement of magnetic field intensity in the chamber.

Stereological analysis of megakaryocytes population

Newborn pups ($n=180$) were weighed. Their spleens were removed and fixed in 10% formalin. Tissue sections of spleens were prepared according to stereological methods and stained with H&E¹⁷⁻¹⁹.

Cavalieri method was used for the numerical evaluation of newborn spleen megakaryocytes¹⁷. Briefly, 30-40 microscopic fields were selected in each section via a systematic subsample. The number of the megakaryocytes was estimated using "dissector" method at the final magnification of 3500 by means of counting frame with inclusion (right and upper) and exclusion (left and lower) borders that was superimposed on the images¹⁷⁻¹⁹.

To estimate the volume of spleens, 15 double serial tissue sections of each sample were selected and stained¹¹⁻¹³. The stereological probe (consisting of 25 points) was superimposed upon the images of the tissue sections viewed on the computer monitor. To take advantage of the probe, for each couple of

sections, two randomly chosen images belonging to the two sections were projected over a computer monitor, one as reference and the other to look up¹⁷⁻¹⁹. The volume of spleen was estimated using the formula:

$$V_v = P_{\text{structure}}/P_{\text{reference}}$$

where 'Pstructure' and 'Preference' are the number of test points falling on the structure's profile and on the reference space, respectively.

To gain the proper estimation of the volume of spleens and the number of cells, five microscopic fields were chosen from each section, projected and counted.

The data are reported as means \pm SD. Statistical analysis was done by One-Way Analysis of Variance (ANOVA) using SPSS 16.0 for Windows XP (SPSS Inc., Chicago, IL). *P* value < 0.05 was considered significant.

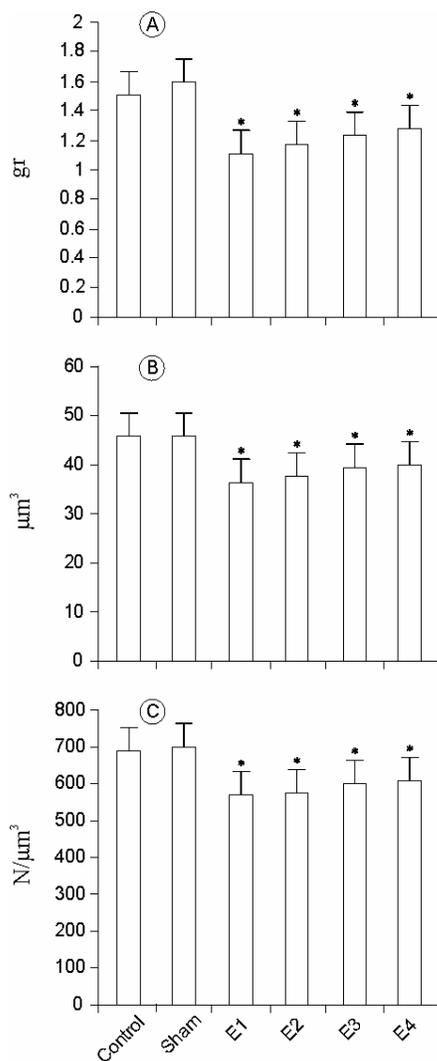
Results

The mean weight of the newborns decreased in ELF-MF exposed groups (E1-E4) and the change was higher in E1 group (treatment at 1-5 days) ($P < 0.024$) (Fig. 1A). The mean volume of the spleens and megakaryocytes number of the newborns decreased and this reduction was more prominent for E1 group (Fig. 1B and C). The mean volume of the spleens and the number of megakaryocytes of the newborns were calculated by stereological method based on tissue section samples (Fig. 2).

Discussion

The results showed that ELF-MF disturbed the development of spleen by decreasing the spleen volume, and the number of megakaryocytes.

There are some reports about the effects of ELF-MF on the embryonic development of spleen and lymphoid organs. Shafey *et al.*²⁰ have shown that exposure to ELF-MF (60 Hz) during chicken pregnancy increased the weight of spleen, lymphoid organs and total body of newborns. However, post-natal birds at 42nd day of age had significantly ($P < 0.01$) lower total body weight, lymphoid organ, and bursa of Fabricius weight when compared with those of 21st day of age, suggesting that although the weight of spleen increased during first days of postnatal life following prenatal exposure to ELF-MF, it reduced later to normal size²⁰. Moreover, exposure to ELF-MF (60 Hz) during 18-21 days of gestation



did not promote inflammatory reactions such as gross or microscopic changes in the spleen²¹.

Another study showed that the volume of spleen in exposed newborn animals was reduced. This result was in contrast with that of Cicekcibasi *et al.*²² who showed that 50 Hz magnetic fields did not reduce the weight of spleen in adult mice²².

In present study the number of splenic megakaryocytes of newborn mice was reduced by prenatal exposure to ELF-MF. The *in vivo* studies have showed some controversy with this data. Gorczyńska²³ showed that static magnetic field (0.05 and 0.3 T) decreases the megakaryocytes in bone-marrow of guinea pigs. But in contrast, Prisco *et al.*²⁴ showed that 900 MHz ELF-MF did not affect the bonemarrow precursor cells of mice. However, these two studies were undertaken on adult samples and could not find any previous data for embryo exactly for the spleen.

Although the mechanisms by which ELF-MF induce cytotoxicity is not completely understood, apoptosis is considered as main cellular event by many authors. *In vivo* studies showed that 50 Hz 0.2, 3.2 or 6.4 mT for 2 weeks or 4 weeks blocked the cell cycle in S phase in testes of mice²⁵ and 60 Hz 14 mT magnetic field for 8 weeks induced apoptosis in testicular germ cell in mice¹².

In vitro studies confirmed *in vivo* experiments that showed induction of apoptosis in K562 human leukemia cell line by ELF-MF (1 mT, 50 Hz)²⁶, in

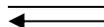


Fig. 1—Effect of maternal exposure to low electro-magnetic fields on (A) body weight (g), (B) spleen volume (μm^3) and (C) number of megakaryocytes ($\text{N}/\mu\text{m}^3$) in newborn mice (* $P < 0.05$).

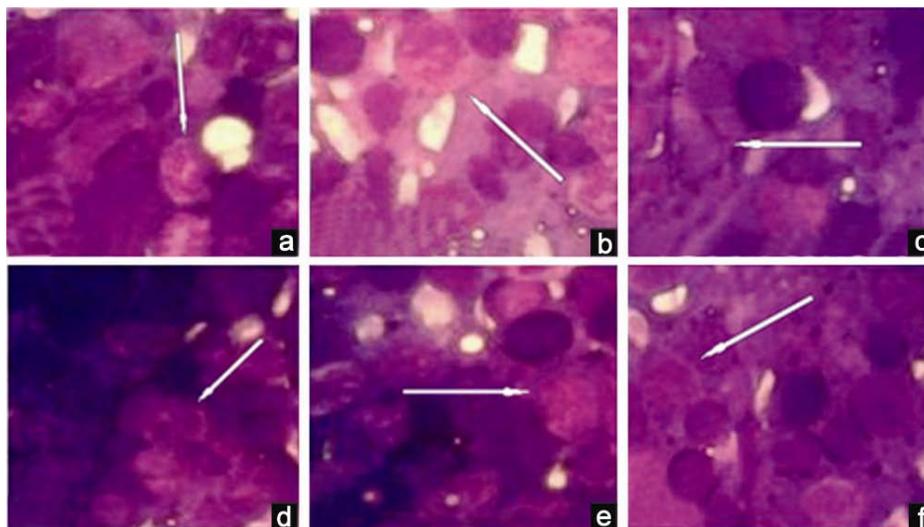


Fig. 2—Effect of maternal exposure to low electro-magnetic fields on spleen of newborn mice [a=sham, b=control, c=E1, d=E2, e=E3, f=E4. (H&E $\times 40$). The arrows show the megakaryocytes.

isolated human osteoclasts by pulsed electromagnetic fields (7.5 Hz)²⁷, in isolated liver cells and peripheral blood sampled from newborn mice by ELF (50 Hz)²⁸, in human normal and cancer cells by time-varying magnetic fields (MFs) (60-Hz 6 mT)²⁹, and also induction of micronuclei in rat bonemarrow by (910-MHz)³⁰. In sub cellular level, regulations of many molecules were considered for induction of apoptosis. The studies suggest formation of free radicals such as reactive oxygen species (ROS)^{26,31}, heat-shock protein 70 (hsp70)^{26,32}, cyclic adenosine mono phosphate (cAMP)³¹ and also P53 gene³³⁻³⁵ and its related gene; P38²⁴ and other regulating genes of apoptosis such as bcl- 2 and bax³³. But an *in vivo* experiment found that these fields had no effect on the expression of P53 gene in testes of rats³⁰ and also an *in vitro* experiment showed that these fields did not inhibit cell proliferation of isolated mouse bonemarrow mesenchymal stem cells³¹. Thus, decreased number of megakaryocytes of newborn mice in ELF-MF exposed groups can be explained by apoptosis process and complete data could be achieved by evaluating ROS and expression of P53 and other related genes.

The present study was conducted to show the effect of 5 days exposure of ELF-MF (50 Hz) during gestational period. Exposure to 900 MHz electromagnetic fields during total time of gestation induced pyramidal cell loss in the hippocampus of newborn rats³⁶. On the other hand, Okudan *et al.*³⁷ have shown that both the static and 50 Hz EFs influence the early development of rat bones at exposure time in 14 days before and after their birth. In fact, duration of exposure during gestational period is more prominent than the frequency and other physical parameters of ELF-MF. The present results also confirm that exposure to ELF-MF during 5 days may induce side-effects like exposure during total period of gestation. These findings are in agreement with those showing that exposure for 1 day to ELF-MF lead to increase cAMP but 2 or 3 days decreases the concentration of this intracellular secondary messenger³¹.

The present results showed that the weights of newborns in all experimental groups were reduced; demonstrating that the whole gestational period is sensitive to these fields for weight loss of delivered newborns. These results are in parallel with the study done on rats subjected to a 35 mW/cm² ELF-MF³⁸ and with the one on mice treated with 20 kHz fields³⁹, but not in agreement with the results on rats treated with 20 kHz fields⁴⁰. The reduction of the weight of

newborn mice may be justified with induction of apoptosis by ELF-MF like the one explained for decreasing the number of megakaryocytes.

In conclusion, this study showed that ELF-MF has a general cytotoxic effect on the total body weight of embryo. These fields may reduce the function of hematopoietic organs, as indicated by reduction in spleen volume and number of megakaryocytes that can be explained by occurrence of cell death. Taking together, using ELF-MF producing instruments like mobile phones and microwave ovens should be limited during gestation. The first trimester of gestation is important because the side-effects of these fields on the embryos are prominent and the mothers usually are unaware of pregnancy.

Acknowledgement

Thanks are due to Arak University of Medical Sciences for financial assistance by a research grant (no. U- 60) and to the staff of Department of Physics for assistance in ELF-MF producing system.

References

- 1 Ahlbom A, Feychting M, Green A, Kheifets L, Savitz DA & Swerdlow AJ, Epidemiologic evidence on mobile phones and tumor risk: A review, *Epidemiology*, 20 (2009) 639.
- 2 Juutilainen J, Matilainen P, Saarikoski S, Laara E & Suonio S, Early pregnancy loss and exposure to 50-Hz magnetic fields, *Bioelectromagnetics*, 14 (1993) 229.
- 3 Lindbohm ML, Hietanen M, Kyrrönen P, Sallmén M, von Nandelstadh P, Taskinen H, Pekkarinen M, Ylikoski M & Hemminki K, Magnetic fields of video display terminals and spontaneous abortion, *Am J Epidemiol*, 136 (1992) 1041.
- 4 Berman E, Chacon L, House D, Koch BA, Koch WE, Leal J, Løvtrup S, Mantipl E, Martin AH, Martucci GI, Mild KH, Monahan JC, Sandström M, Shamsaifar K, Tell K, Trillo MI, Ubeda A & Wagner P, Development of chicken embryos in a pulsed magnetic field, *Bioelectromagnetics*, 11 (1990) 169.
- 5 Farrell JM, Litovitz TL, Penafiel M, Montrose CJ, Doinov P, Barber M, Brown KM & Litovitz TA, The effect of pulsed and sinusoidal magnetic fields on the morphology of developing chick embryos, *Bioelectromagnetics*, 18 (1997) 431.
- 6 Youbicier-Simo BJ, Boudard F, Cabaner C & Bastide M, Biological effects of continuous exposure of embryos and young chickens to electromagnetic fields emitted by video display units, *Bioelectromagnetic*, 18 (1997) 514.
- 7 Huuskonen H, Lindbohm ML & Juutilainen J, Teratogenic and reproductive effects of low-frequency magnetic fields, *Mutat Res*, 410 (1998) 167.
- 8 Ryan BM, Mallett E Jr, Johnson TR, Gauger JR & McCormick DL, Developmental toxicity study of 60 Hz (power frequency) magnetic fields in rats, *Teratology*, 54 (1996) 73.
- 9 Kowalczyk CI, Robbins L, Thomas JM, Butland BK & Saunders RD, Effects of prenatal exposure to 50 Hz magnetic fields on development in mice, *Bioelectromagnetics*, 15 (1994) 349.

- 10 Dundar B, Cesur G, Comlekci S, Songur A, Gokcimen A, Sahin O, Ulukut O, Yilmaz HR, Sutcu R & Caliskan S, The effect of the prenatal and post-natal long-term exposure to 50 Hz electric field on growth, pubertal development and IGF-1 levels in female Wistar rats, *Toxicol Ind Health*, 25 (2009) 479.
- 11 Dupont MJ, Parker G & Persinger MA, Reduced litter sizes following 48-h of prenatal exposure to 5 nT to 10 nT, 0.5 Hz magnetic fields: implications for sudden infant deaths, *Int J Neurosci*, 115 (2005) 713.
- 12 Kim YW, Kim HS, Lee JS, Kim YJ, Lee SK, Seo JN, Jung KC, Kim N & Gimm YM, Effects of 60 Hz 14 mT magnetic field on the apoptosis of testicular germ cell in mice, *Bioelectromagnetics*, 30 (2009) 66.
- 13 Lukac T, Matavulj A, Matavulj M, Rajković V & Lazetić B. Photoperiodism as a modifier of effect of extremely low-frequency electromagnetic field on morphological properties of pineal gland, *Bosn J Basic Med Sci*, 6 (2006) 10.
- 14 Odaci E, Bas O & Kaplan S. Effects of prenatal exposure to a 900 MHz electromagnetic field on the dentate gyrus of rats: A stereological and histopathological study, *Brain Res*, 1238 (2008) 224.
- 15 Reipert BM, Allan D, Reipert S & Dexter TM. Apoptosis in haemopoietic progenitor cells exposed to extremely low-frequency magnetic fields, *Life Sci*, 61 (1997) 1571.
- 16 Dawson TW, Caputa K, Stuchly MA & Kavet R, Comparison of electric fields induced in humans and rodents by 60-Hz contact currents, *IEEE Trans Biomed Eng*, 50 (2003) 744.
- 17 Howard CV & Reed MG, *Three-dimensional measurement in microscopy*. (Oxford University Press, London) 1998, 20.
- 18 Karbalay-Doust S & Noorafshan A, Stereological study of the effects of nandrolone decanoate on the mouse liver, *Micron*, 40 (2009) 471.
- 19 Noorafshan A, Esmail-Zadeh B, Bahmanpour S & Poost-Pasand H, Early stereological changes in liver of Sprague-Dawley rats after streptozotocin injection, *Indian J Gastroenterol*, 24 (2005) 104.
- 20 Shafey TM, Al-Mufarej S & Al-Batshan HA, Effect of electric field during incubation of eggs on the immune responses of hatched chickens, *Electromagn Biol Med*, 25 (2006) 163.
- 21 Mandeville R, Franco E, Sidrac-Ghali S, Paris-Nadon L, Rocheleau N, Mercier G, Désy M, Devaux C & Gaboury L, Evaluation of the potential promoting effect of 60 Hz magnetic fields on N-ethyl-N-nitrosourea induced neurogenic tumors in female F344 rats, *Bioelectromagnetics*, 21 (2000) 84.
- 22 Cicekcibasi AE, Celik I, Salbacak A, Ozkan Y, Okudan N & Buyukmumcu M, Determination of the effects of extremely low frequency electromagnetic fields on the percentages of peripheral blood leukocytes and histology of lymphoid organs of the mouse, *Saudi Med J*, 29 (2008) 36.
- 23 Gorczyńska E, The process of myelopoiesis in guinea pigs under conditions of a static magnetic field, *Acta Physiol Pol*, 38 (1987) 425.
- 24 Prisco MG, Nasta F, Rosado MM, Lovisolio GA, Marino C & Pioli C, Effects of GSM-modulated radiofrequency electromagnetic fields on mouse bonemarrow cells, *Radiat Res*, 170 (2008) 803.
- 25 Hong R, Liu Y, Yu YM, Hu K & Weng EQ, Effects of extremely low frequency electromagnetic fields on male reproduction in mice, *Zhonghua Lao Dong Wei Sheng Zhi Ye Bing Za Zhi*, 21 (2003) 342.
- 26 Garip AI & Akan Z, Effect of ELF-EMF on number of apoptotic cells; correlation with reactive oxygen species and HSP, *Acta Biol Hung*, 61 (2010) 158.
- 27 Chang K, Chang WH, Tsai MT & Shih C, Pulsed electromagnetic fields accelerate apoptotic rate in osteoclasts, *Connect Tissue Res*, 47 (2006) 222.
- 28 Udroui I, Cristaldi M, Ieradi LA, Bedini A, Giuliani L & Tanzarella C, Clastogenicity and aneuploidy in newborn and adult mice exposed to 50 Hz magnetic fields, *Int J Radiat Biol*, 82 (2006) 561.
- 29 Kim J, Ha CS, Lee HJ & Song K, Repetitive exposure to a 60-Hz time-varying magnetic field induces DNA double-strand breaks and apoptosis in human cells, *Biochem Biophys Res Commun*, 400 (2010) 739.
- 30 Demsia G, Vlastos D & Matthopoulos DP, Effect of 910-MHz electromagnetic field on rat bonemarrow, *Scient World J*, 4 (2004) 48.
- 31 Wu H, Ren K, Zhao W, Baojian GE & Peng S, Effect of electromagnetic fields on proliferation and differentiation of cultured mouse bonemarrow mesenchymal stem cells, *J Huazhong Univ Sci Technolog Med Sci*, 25 (2005) 185.
- 32 Simkó M, Droste S, Kriehuber R & Weiss DG, Stimulation of phagocytosis and free radical production in murine macrophages by 50 Hz electromagnetic fields, *Eur J Cell Biol*, 80 (2001) 562.
- 33 Tenuzzo B, Vergallo C & Dini L, Effect of 6 mT static magnetic field on the bcl-2, bax, p53 and hsp70 expression in freshly isolated and *in vitro* aged human lymphocytes, *Tissue Cell*, 41 (2009) 169.
- 34 Hasegawa M, Zhang Y, Niibe H, Terry NHA & Meistrich ML, Resistance of differentiating spermatogonia to radiation induced apoptosis and loss in p53-deficient mice, *Radiat Res*, 149 (1998) 263.
- 35 Akdag MZ, Dasdag S, Aksen F, Isik B & Yilmaz F, Effect of ELF magnetic fields on lipid peroxidation, sperm count, p53, and trace elements, *Med Sci Monit*, 12 (2006) 66.
- 36 Bas O, Odaci E, Mollaoglu H, Ucok K & Kaplan S, Chronic prenatal exposure to the 900 megahertz electromagnetic field induces pyramidal cell loss in the hippocampus of newborn rats, *Toxicol Ind Health*, 25 (2009) 377.
- 37 Okudan B, Keskin AU, Aydin MA, Cesur G, Cömlekçi S & Süslü H, DEXA analysis on the bones of rats exposed *in utero* and neonatally to static and 50 Hz electric fields, *Bioelectromagnetics*, 27 (2006) 589.
- 38 Jensh RP, Behavioral teratologic studies using microwave radiation: is there an increased risk from exposure to cellular phones and microwave ovens? *Reprod Toxicol*, 11 (1997) 601.
- 39 Svedenstål BM & Johanson KJ, Fetal loss in mice exposed to magnetic fields during early pregnancy, *Bioelectromagnetics*, 16 (1995) 284.
- 40 Lee HJ, Pack JK, Gimm YM, Choi HD, Kim N, Kim SH & Lee YS, Teratological evaluation of mouse fetuses exposed to a 20 kHz EMF, *Bioelectromagnetics*, 30 (2009) 330.